



Evaluation of antimicrobial effects of *Zataria multiflora*, *Rosemarinus officinalis* and *Cuminum cyminum* essential oils on the growth of *Escherichia coli* O157:H7

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ABSTRACT

Due to the increased resistance to antibiotics, identifying new compounds with fewer side effects is more important than usual medications in treatment of microorganisms. This study aimed to examine the antibacterial activity of *Zataria multiflora*, *Rosemarinus officinalis* and *Cuminum cyminum* essential oils on the growth of *E. coli* O157:H7 as one of the most common pathogens of food-borne diseases. *Z. multiflora*, *R. officinalis* and *C. cyminum* plants were dried and ground. Then the extraction of essential oil was performed by the hydrodistillation method. In order to examine the antibacterial effects, micro-broth dilution was used to determine the minimum inhibitory concentration (MIC) and minimum bactericidal concentrations (MBC). The disc diffusion method was applied to measure the inhibitory zone diameter. The data were analyzed by SPSS. Maximum diameter of inhibition zone (26.96 mm) was related to *Z. multiflora* which had a significant difference compared to other essential oils ($p < 0.001$). Also the MIC and MBC of *Z. multiflora* essential oil were obtained as 3.125 mg/ml and 6.25 mg/ml, respectively. In this study, *Zataria multiflora*, *Rosemarinus officinalis* and *Cuminum cyminum* essential oils had significant antibacterial properties against *E. coli* O157:H7. As a result of these essential oils after completing studies, they can be a good alternative to chemical preservatives for storage of food.

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1. Introduction

Over the past decades, food borne diseases are considered as one of the leading public health threats worldwide. These diseases are classified as intestinal diseases and have been increasing, not only in developing countries, but also in developed countries (1,3). Various factors contribute to the transmission of food-borne diseases including bacteria, viruses, parasites, fungi and some toxins (4,5) that bacteria and toxins derived from them are the most common cause of food-borne diseases (6,7).

E. coli O157:H7 is one of the most important

serotypes of enterohemorrhagic *Escherichia coli* (EHEC) that due to low infectious dose (less than 100 pcs), these bacteria are transmitted to human by consumption of contaminated food (8,9). STX1 and STX2 are the most important toxins in these bacteria that with other bacteria cause some disease such as hemolytic uremic syndrome (HUS), hemorrhagic colitic and thrombotic thrombocytopenic purpura (12,16).

Studies have shown that various extracts and essential oils prevent the growth of a wide range of pathogenic microorganisms. A reason for this approach can be seen in the presence of natural origin and the lack of adverse effects in humans and in the environment as to chemical preservatives (17).

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Examples of medicinal herbs with their antimicrobial effects are *Z. multiflora*, *R. officinalis*, and *C. cyminum*. *Z. multiflora* is the most well-known medicinal herb which grows in Iran, Afghanistan and Pakistan. This herb is known as an antiseptic, anti-spasm and anti-inflammatory agent in traditional medicine. Also, *Z. multiflora* is widely used as a food flavoring that is related to phenolic compounds such as caracrole, eugenol and thymol in this herb (18,19).

R. officinalis, commonly known as rosemary, is widely known as a spice and used in traditional medicine (20,21). It is a member of the mint family Lamiaceae and is native to Asia and some other parts of the world. Also, *C. cyminum* can be used in the treatment of digestive and pulmonary diseases (22,23).

Considering a wide range of medicinal herbs in Iran, the present study was conducted to examine the effect of the concentrated essential oils of *Z. multiflora*, *R. officinalis* and *C. cyminum* on the growth rate of *E.coli* O157: H7.

2. Materials and methods

2.1. Preparation of essential oil

For essential oil preparation, plants of *Z. multiflora*, *R. officinalis* and *C. cyminum* were obtained in spring of 2016 from Maragheh and Tabriz cities, Iran. For the identification, these plants were sent to Department of Plant Biology of Payame Noor University of Maragheh. The plants were dried in shading conditions and 50 g of them were individually powdered by electric grinders and added to a balloon containing 700 ml of distilled water and were prepared for Hydrodistillation. The essential oils were dewatered and dried using Sodium Sulfate and Whatman Filter (Whatman 2) and kept in dark-colored containers away from light and kept in a fridge (24).

2.2. Bacterial strains

For evaluation of the antibacterial effect of essential oil, the standard strain of *E. coli* O157: H7 NCTC12900 was selected. For fresh culturing of isolates, nutrient broth medium (Merck, Germany) was used and to provide a uniform bacterial suspension, after 24 hours of culturing of strains, 2-3 colonies were inoculated into 5 ml sterile normal saline and placed at 37°C for 30 minutes until the opacity was equivalent to 0.5 McFarland standard (1.5×10^8 cfu/ml) (25).

2.3. Antibacterial activity of the essential oil

Antimicrobial properties of the essential oils were determined using the disc diffusion method and based on Clinical and Laboratory Standards Institute Protocols (CLSI) (26). The essential oils were solved in dimethyl sulfoxide (DMSO) solvent and after preparation, concentrations of 200, 100, 50, 25, 12.5, 6.25, 3.125 and 1.562 mg/ml were sterilized by passing through a 0.22 μ m pore size filter. After culturing of bacterial strain on Mueller-Hinton agar plates, 6 mm paper discs containing the essential oils with concentrations of 10 μ l were placed on plate surface and were incubated at 37°C for 18-24 hours. Enrofloxacin (5 μ g), flumequin (30 μ g), oxytracycline (30 μ g) and erythromycin (15 μ g), were used for antibiotic susceptibility testing and discs containing DMSO were used as negative control. All experiments were performed three times (25).

2.4. Minimum Inhibitory Concentration (MIC)

Micro-broth dilution method was used to determine the minimum inhibitory concentration, according to the CLSI protocol (26). Briefly, 20 μ l of the essential oils dissolved in DMSO was added to each 96-well microplate well containing 160 μ l Mueller-Hinton broth (Merck, Germany) and serial dilutions were provided. Then, 20 μ l of microbial suspension containing 10^6 cfu/ml was added to each well of the microplates. The negative and positive controls of the culture medium and bacterial suspension were added to the wells. Then all microplates were incubated at 37°C for 18-24 h.

2.5. Minimum Bactericidal Concentration (MBC)

For this section, 5 μ l suspension from all wells without turbidity in the MIC phase were cultured on the Mueller-Hinton agar medium. Afterwards, the plates were incubated at 37°C for 24 h. The lowest concentration of essential oil in which the bacteria were killed was regarded as MBC. The mean values of inhibition zones were compared using one-way ANOVA and SPSS 23.

3. Results

The results of this study revealed that essential oils of *Z. multiflora*, *R. officinalis* and *C. cyminum* can affect the growth of *E.coli* O157:H7. The lowest inhibitory levels of growth were observed at 10 mg/ml of *Z. multiflora* and *R. officinalis*. *Z. multiflora* had the highest effect on *E.coli* O157: H7 (with 26.96 mm diameter of the inhibition zones) that in this regard, a significant

difference was observed between the other essential oils ($p < 0.001$). Also, *Z. multiflora* with MIC= 3.125 and MBC=6.25 mg/ml had an antimicrobial activity equivalent to tetracycline and much more than erythromycin. The results of antimicrobial activity of essential oils and antibiotics are shown in tables 1 and 2.

Table 1. The mean diameter of the inhibition zones in the standard strain of *E.coli* O157: H7 (the concentration of 10 mg/ml)

Essential oil	The average diameter of the inhibition zones (mm)
<i>Zataria multiflora</i>	26.96
<i>Rosemarinus officinalis</i>	14
<i>Cuminum cyminum</i>	-

Table 2. The MIC and MBC of antibiotics and essential oils against the standard strain of *E.coli* O157: H7

Antibiotic	(MIC) (mg/ml)	(MBC) (mg/ml)
Enrofloxacin	0.781	1.562
Flumequin	1.562	3.125
Oxytracycline	3.125	6.25
Erythromycin	6.25	12.5
<i>Zataria multiflora</i>	3.125	6.25
<i>Rosemarinus officinalis</i>	12.5	25
<i>Cuminum cyminum</i>	25	50

4. Discussion

The findings of this study showed that *Z. multiflora*, *R. officinalis* and *C. cyminum* essential oil have antimicrobial effect on *E.coli* O157:H7. These bacteria are one of the most important pathogenic agents of gastrointestinal diseases that can lead to high mortality during epidemics and outbreaks due to the consumption of contaminated foods (13). Hence, one of the main concerns of food industry owners and consumers of food is contamination with *E.coli* O157:H7 and other microbial agents. In many cases, antibiotics and chemical preservatives are used to boost the shelf-life of foods and block the pathogenic bacteria's growth or reproduction. The growth of bacteria can lead to food corrosion that causes a lot of public health problems. One of the problems is increasing of resistance to antibiotics in food-borne bacteria. Therefore, one of methods for controlling of food pathogens is the use of the plant-derived compounds (28,29).

In recent years, pharmaceutical companies have carried out extensive research on the discovery of safe and effective antimicrobial agents for the prevention and treatment of a wide range of multi-drug resistance bacterial infections. Because of antimicrobial effects of the essential oils and their lower side effects, these agents can be used as an appropriate alternative to antibiotics (30). The results of this study evinced that

the essential oils of *Z. multiflora* and *R. officinalis* can inhibit the growth of *E. coli* O157: H7. As the concentration of the extracts increased, the inhibition zone increased and the highest inhibitory effect of the bacteria was observed at a concentration of 10 mg/ml. Also, the essential oil of *Z. multiflora* was significantly more effective than other essential oils ($p < 0.001$) and its antimicrobial effect was much stronger than erythromycin. In recent years, the antimicrobial effects of the extracts and essential oils have been reported (31).

Soltan Dallal et al. studied the effect of *Z. multiflora* on antibiotic resistant *Staphylococcus aureus* strains isolated from the food. They reported that these agents significantly can reduce the growth of *S. aureus* (34). Also, Yaghobzadeh et al. in 2011 investigated the effect of *Z. multiflora* on shiga-toxin producing *E. coli* strains. In this study, the mean diameter of the bacterial inhibition zone was 28.18 mm, which is in line with the findings of the current study (35). In relation to the antibacterial effects of *R. officinalis*, Soltan Dallal et al. showed that the diameter of the inhibition zone was observed about 20 mm in methicillin-resistant staphylococci isolated from patients and foodstuffs (30). In another study by Malakootian and Hatami, the diameter of the inhibition zone was 16 ± 1.07 mm, which is similar to the findings of the current study. It was also found that the MIC from the growth of the *E. coli* was 3000 $\mu\text{g/ml}$ and its MBC was 3200 $\mu\text{g/ml}$ (25).

Also, studies have done on the antimicrobial effect of *C. cyminum* (23,36,37,38). According to the findings of the present study, the essential oil of *C. cyminum* had a very low inhibitory effect and less than < 1 mg/ml on the *E. coli* O157: H7. This could be due to the concentration of essential oils tested in this study, because Suliman in Iraq (2009) showed that 400 mg/ml of methanolic extract of *C. cyminum* have inhibitory effect on the growth of Gram-negative bacteria (39). Similarly to the results of this study, Haghighi et al. reported that *C. cyminum* in comparison with other essential oils had the lowest antifungal effect on *Candida albicans* (40).

Due to the many antimicrobial effects of the essential oils, they can be used as natural preservatives to increase the shelf-life of foods. The limitations of the present study are not to use concentrations higher than 10 mg/ml because the use of high concentrations of extracts and essential oils, in some cases, causes changes in the organoleptic properties of foods (31). According to the results of this study, it is possible to develop appropriate drugs for the elimination of food-borne bacteria by using these essential oils.

5. Conclusion

This study showed that essential oil of *Z. multiflora* and *R. officinalis* have antimicrobial effects against *E. coli* O157: H7, which can be used as an appropriate herbal source against microorganisms and treatment of human bacterial pathogens.

Conflict of interest

The authors declared that they have no conflict of interest.

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References

- Burt S. Essential oils: Their Antibacterial Properties and Potential Applications in foods, a Review. *Int J Food Microbiol* 2004; 94: 223-53.
- Garcia P, Rodriguez L, Rodriguez A, et al. Food biopreservation: promising strategies using bacteriocins, bacteriophages and endolysins. *Trends Food Sci Technol* 2010; 8: 373-82.
- Badrei N, Gobin A, Dookeran S, et al. Consumer awareness and perception to food safety hazards in Trinidad, West Indies. *Food Control* 2006; 17: 370-77.
- Wang XW, Zhang L, Jin LQ, et al. Development and application of an oligonucleotide microarray for the detection of foodborne bacterial pathogens. *Appl Microbiol Biotechnol* 2007; 76: 225-33.
- Sarshar M, Doosti A, Jafari A, et al. Evaluation of oligonucleotide microarray technology for the detection of foodborne bacterial pathogens. *J Microbial World* 2009; 2: 73-80 [in persian].
- Headrick ML, Tollefson L. Food borne disease summary by food commodity. *Vet Clin North Americ Food Animal Pract* 1998; 14: 91-100.
- Baldursson S, Karanis P. Waterborne transmission of protozoan parasites: review of worldwide outbreaks—an update 2004-2010. *Water Res* 2011; 45: 6603-14.
- Momose Y, Hirayama K, Iton K. Effect of organic acids on inhibition of *Escherichia coli* O157:H7 colonization in gnotobiotic mice associated with infant intestinal microbiota. *Antonie Leeu Wenhoek* 2008; 93:141-49.
- Chaucheyras-Durand F, Madic J, Dondin F, et al. Biotic and abiotic factors influencing in vitro growth of *Escherichia coli* O157:H7 in ruminant digestive contents. *Appl Environ Microbiol* 2006; 72: 4136-42.
- Tavakoli HR, Sarraf pour R, Samadi M. Water, Food and Bioterrorism. *J Mil Med* 2005; 7: 75-82 [in persian].
- Centers for Disease Control and Prevention. 2008-2012. Retrieved 2009-05-22.
- Wang G, Clark CG, Rodgers FG. Detection in *Escherichia coli* of the genes encoding the major virulence factors, the genes defining the O157:H7 serotype, and components of the Type 2 Shiga toxin family by multiplex PCR. *J Clin Microbiol* 2002; 40: 3613-19.
- Mehdizadeh M, Eskandari S, Zavar M, et al. The Importance of *Escherichia coli* O157:H7 in Foodborne Infection. *J. Kerman Univ Med Sci* 2008; 15: 353-61 [in persian].
- Jamshidi A, Bassami MR, Rasooli M. Isolation of *Escherichia coli* O157:H7 from ground beef samples collected from beef markets, using conventional culture and polymerase chain reaction in Mashhad, northeastern Iran. *Iran J Vet Res* 2008; 9: 72-76.
- Takahashi M, Taguchi H, Yamaguchi H, et al. The effect of probiotic treatment with *Clostridium butyricum* on enterohemorrhagic *Escherichia coli* O157:H7 infection in mice. *FEMS Immunol Med Microbiol* 2004; 41:219-26.
- Shekarforoush SS, Nazer AHK, Firouzi, R et al. Effects of storage temperatures and essential oils of oregano and nutmeg on the growth and survival of *Escherichia coli* O157:H7 in barbecued chicken used in Iran. *J Food Control* 2007; 18: 1428-33.
- AAla F, Khodaveysi S, Baghdadi E, et al. The antifungal effect of *Pistacia Atlantica* subsp. *Kurdica* on the growth of *Aspergillus parasiticus*. *J Res Health Sci* 2015; 13: 79-84 [in persian].
- Hashemi A, Shams S, Barati M, et al. Antibacterial effects of metabolic extracts of *Zataria multiflora*, *Myrtus communis* and *peganum harmala* on *Pseudomonas aeruginosa* producing ESBL. *Arak Med Uni J* 2011; 14: 104-12 [in persian].
- Burt S. Essential oils: their antibacterial properties and potential applications in foods—a review. *Int J Food Microbiol* 2004; 94: 223-53.
- Campo JD, Amiot MJ, Nguyen-The C. Antimicrobial effect of rosemary extracts. *J Food Protect* 2000; 63: 1359-68.
- Ozcan M. Antioxidant activities of rosemary, sage, and sumac extracts and their combinations on stability of natural peanut oil. *J Med Food* 2003; 6: 267-70.
- Oroojalian F, Kasra-Kermanshahi R, Azizi MA, et al. Synergistic antibacterial activity of the essential oils from three medicinal plants against some important food-borne pathogens by microdilution method. *Iranian J Med & Arom Plant* 2010; 26: 133-46.
- Gachkar L, Yadegari D, Rezaei MB, et al. Chemical and biological characteristics of *Cuminum cyminum* and *Rosmarinus officinalis* essential oils. *Food Chem* 2007; 102: 898-04.
- Mohammadi KH, Karim G, Hanifian SH, et al. Antimicrobial effect of *Zataria multiflora* Boiss. essential oil on *Escherichia coli* O157:H7 during manufacture and ripening of white brined cheese. *J Food Hyg* 2011; 1: 69-78 [in persian].
- Malakootian M, Hatami B. Survey of Chemical Composition and Antibacterial Activity of *Rosmarinus Officinalis* Essential oils on *Escherichia Coli* and Its Kinetic. *J Toloo-e-behdasht* 2013; 12: 1-13 [in persian].

26. CLSI. Performance standards for antimicrobial susceptibility testing; 24th informational supplement. CLSI document M100-S24. Wayne, PA: Clinical and Laboratory Standards Institute; 2014.
27. Demirci F, Guven K, Demirci B, et al. Antibacterial activity of two *Phlomis* essential oils against food pathogens. *Food Control* 2008; 19: 1159-64.
28. Valero M, Salmeron M. Antibacterial activity of 11 essential oils against *Bacillus cereus* in tyndallized carrot broth. *Int J Food Microbiol* 2003; 85: 73-81.
29. Vernozy-Rozand C, Ray-Gueniot S, Ragot C et al. Prevalence of *Escherichia coli* O157: H7 in industrial minced beef. *Letter Appl Microbiol* 2002; 35: 7-11.
30. Dallal MM, Mashkani MG, Yazdi MH, et al. Antibacterial effects of *Rosmarinus officinalis* on Methicillin-resistant *Staphylococcus aureus* isolated from patients and foods. *Sci J Kurdistan Uni Med Sci* 2011; 16: 73-80 [in persian].
31. Zandi H, Hajimohammadi B, Amiri Aet, al. Antibacterial Effects of (*Mentha X Piperita L.*) Hydroalcoholic Extract on the Six Food-Borne Pathogenic Bacteria. *Toloo-e-behdasht* 2016 ;15: 22-33 [in persian].
32. Ataee M, Hosseini H, Noori N, et al. Effect of *Zataria multiflora* boiss. essential oil on growth curve and shigatoxin 2 production of enterohemorrhagic *Escherichia coli* O157: H7. *J Med Plant* 2013; 4: 62-71 [in persian].
33. Masoomi V, Tajik H, Moradi M, et al. Antimicrobial effects of *Zataria multiflora* boiss. essential oil nanoemulsion against *Escherichia coli* O157: H7. *J Urmia Uni Med Sci* 2016; 27: 608-17 [in persian].
34. Dallal MM, Bayat M, Yazdi MH, et al. Antimicrobial effect of *Zataria multiflora* on antibiotic-resistant *Staphylococcus aureus* strains isolated from food. *Scientific J Kurdistan Uni Med Sci* 2012; 17: 21-29 [in persian].
35. Yaghobzadeh N, Ownagh A, Mardani K, et al. Molecular identification, antibiotic resistance profile of shiga toxin-producing *Escherichia coli*(STEC) and antibacterial activity of *Zataria multiflora* boiss and *Carum copticum* essential oil against them. *Urmia Med J* 2011; 22: 262-69 [in persian].
36. Daneshmandi S, Soleimani N, Sattari M, et al. Evaluation of the drug synergistic and antibacterial effects of *cuminum cyminum* essential oil. *Arak Med Uni J* 2010; 13: 75-82 [in persian].
37. Bonyadian M, Karim G. Study of the effect of some volatile oils of herbs (pennyroyal, peppermint, tarragon, caraway seed and Thyme) against *E.coli* and *S.aureus* in broth media. *J Vet Res* 2002; 57: 81-83.
38. Motamedifar M, Hashemizadeh Z, Haghigati M, et al. Evaluation of Antibacterial Effects of Aqueous and Alcoholic Extracts of *Cuminum cyminum L.* Seeds on Some Pathogenic Bacteria. *Sadra Med Sci J* 2017; 3: 21-30 [in persian].
39. Suliman SK. Effect of crude methanolic extract of *Cuminum cyminum* on growth of some types of pathogenic bacteria. *J Tikrit Uni Agri Sci* 2009; 9: 611-15.
40. Haghghi F, Roudbar Mohammadi S, Soleimani N, et al. Evaluation of antifungal activity of essential oils of *Thymus vulgaris*, *Petroselinum Crispum*, *Cuminum cyminum* and *Bunium persicum* on *Candida albicans* in comparison with Fluconazole. *Pathobiol Res* 2011; 14: 29-35 [in persian].